

# Towards a multi-scale model of combination targeted and cytotoxic therapy to evaluate treatment response in HER2+ breast cancer

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## Overview

- Using an established ODE model at the tissue scale for a preclinical model of HER2+ breast cancer undergoing trastuzumab treatment, expand to include cell scale effects for combination trastuzumab-paclitaxel therapy
- Will connect the two scales by coupling drug effects into the growth and reduction terms of the governing equation for tumor volume changes
- An integrated mathematical-experimental approach bridging *in vivo* and *in vitro* experimental data, and therefore multiple scales, may elucidate the potential best strategies for combination therapy for HER2+ breast cancer

## Tissue Scale Model

- Five coupled, ODEs for the longitudinal relationship of vasculature, hypoxia, necrosis, immune infiltration, and tumor growth<sup>3</sup>

$$\begin{aligned} [1] \quad \frac{dT}{dt} &= gT(1 + \rho H) - \mu_T T I \\ [2] \quad \frac{dI}{dt} &= \alpha_V V(1 - I) + \alpha_N N(1 - I) - \mu_I I T \\ [3] \quad \frac{dV}{dt} &= \alpha_T T(1 - V) + \alpha_I I(1 - V) - \mu_V V T \\ [4] \quad \frac{dN}{dt} &= \beta(1 - V)(1 - N) - \mu_N N I \\ [5] \quad \frac{dH}{dt} &= \gamma(\delta - V)H(1 - H) \end{aligned}$$

Table 1: Definitions of model variables

Output	Description
$T$	Tumor volume
$I$	Fraction of immune response in tumor
$V$	Fraction of well-vascularized tumor
$N$	Fraction of necrosis in tumor
$H$	Fraction of hypoxia in tumor

Table 2: Definitions of model parameters

Parameter	Description
$g$	Tumor volume growth rate
$\rho$	Ability of hypoxia to promote tumor growth
$\mu_T$	Rate tumor volume decreases due to immune response
$\alpha_N$	Rate immune response increases due to necrosis
$\alpha_V$	Rate immune response increases due to vasculature
$\mu_I$	Rate immune response decreases per tumor volume
$\alpha_T$	Rate vascularization increases per tumor volume
$\alpha_I$	Rate vascularization increases due to immune response
$\mu_V$	Rate well-vascularized tissue decreases per tumor volume
$\beta$	Rate necrosis increases due to decreased fraction of well-vascularized tumor
$\mu_N$	Rate necrosis decreases due to immune response
$\gamma$	Rate hypoxia increases or decreases due to fraction of well-vascularized tumor
$\delta$	Threshold for hypoxia to increase or decrease due to fraction of well-vascularized tumor

Figure 2: Diagram of the interactions in the model equations between the different biological components

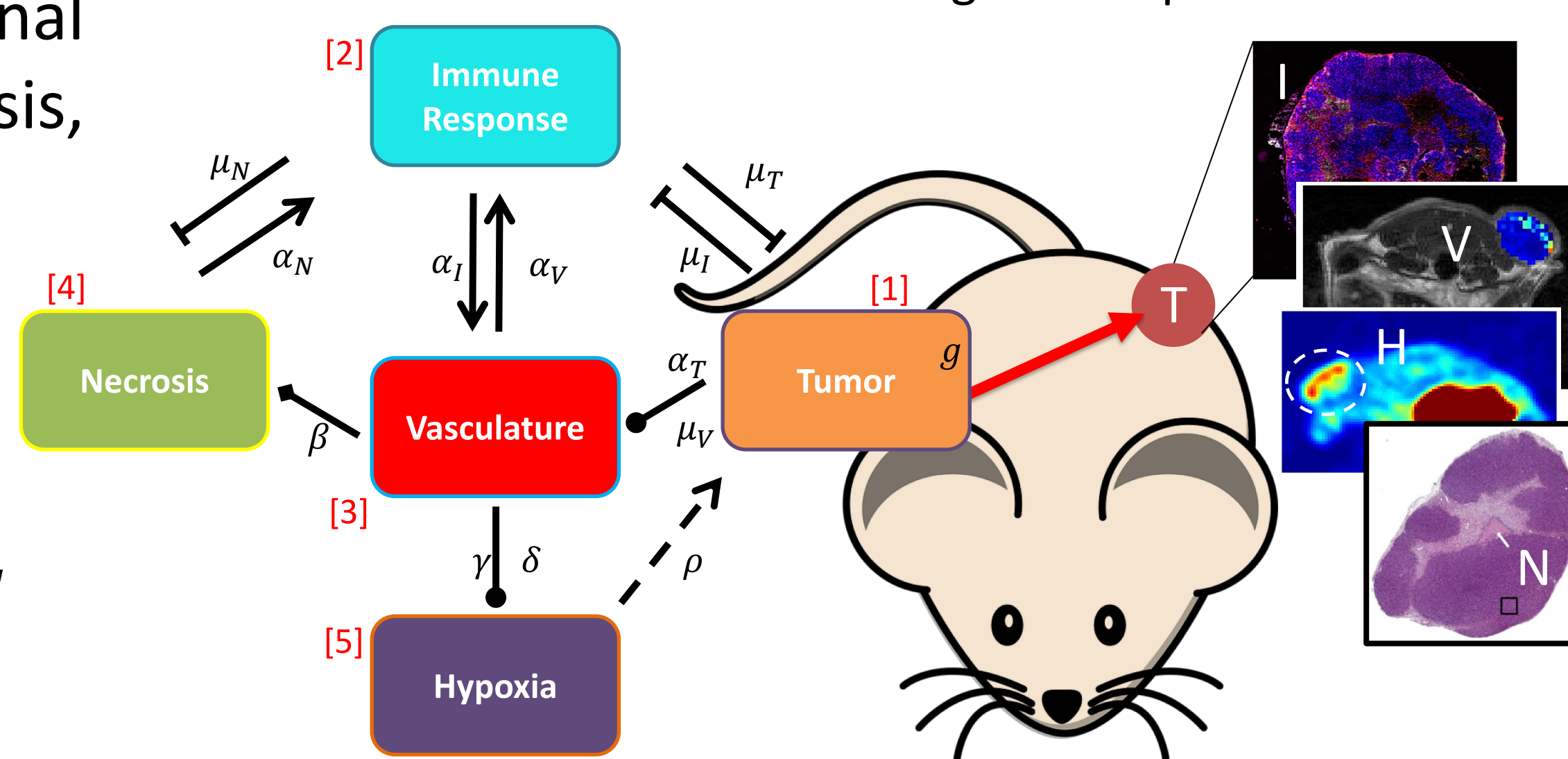
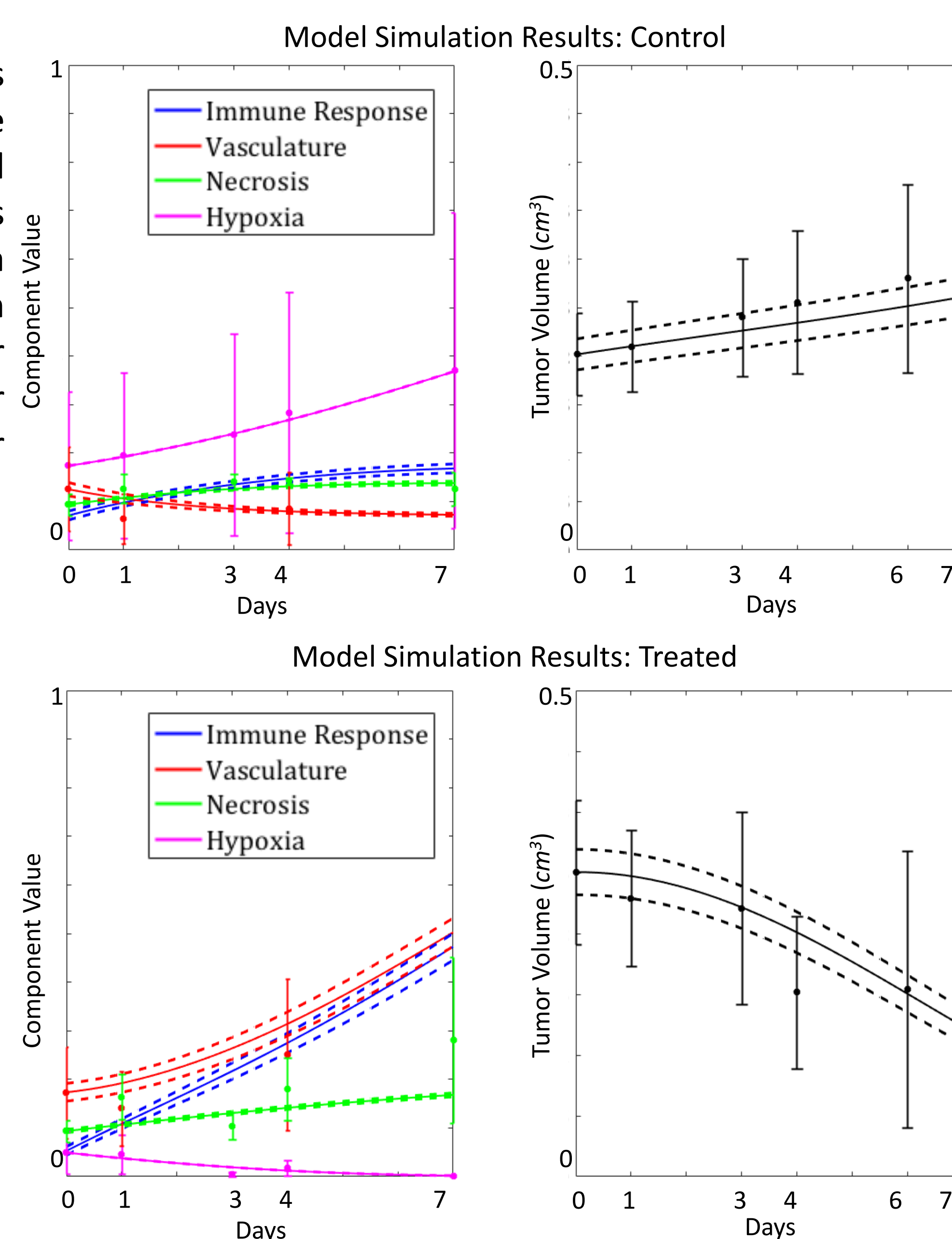


Figure 3: Model results with 95% confidence intervals indicated with dashed lines against validation data for tumor volume with 95% confidence error bars (right: tumor volume, left: tumor tissue components)

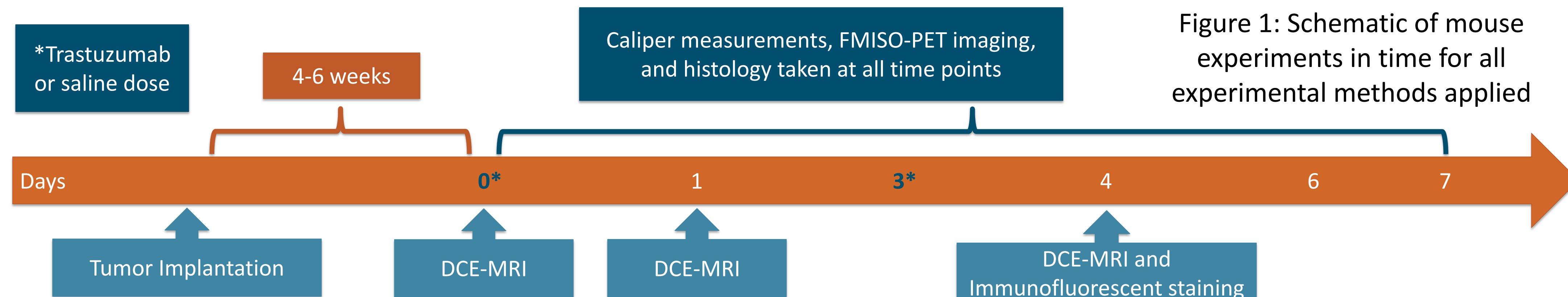


## In vivo data

- Nude athymic mice subcutaneously implanted with BT474 HER2+ human breast cancer cells in the flank (allowed to grow for 4-6 weeks) and injected on days 0 and 3 with trastuzumab (10 mg/kg) or, in controls, with saline.
- Over 7 days, tumor volume, vasculature, necrosis, hypoxia, and immune response measurements quantified using several experimental methods (Table 1)
- Tumor volume collected using caliper measurements
- Physiological parameter for vascular perfusion and permeability,  $K^{trans}$ , derived from dynamic contrast-enhanced magnetic resonance imaging (DCE-MRI) data for fraction of well-vascularized tumor
- Hematoxylin and eosin (H&E) staining for percent necrosis
- <sup>18</sup>F-fluoromisonidazole positron emission tomography (FMISO-PET) standard uptake values for percent hypoxia.
- Immunofluorescent imaging for CD11c & F4/80 myeloid markers for immune infiltration

Table 1: Summary of experimental data utilized and their sources

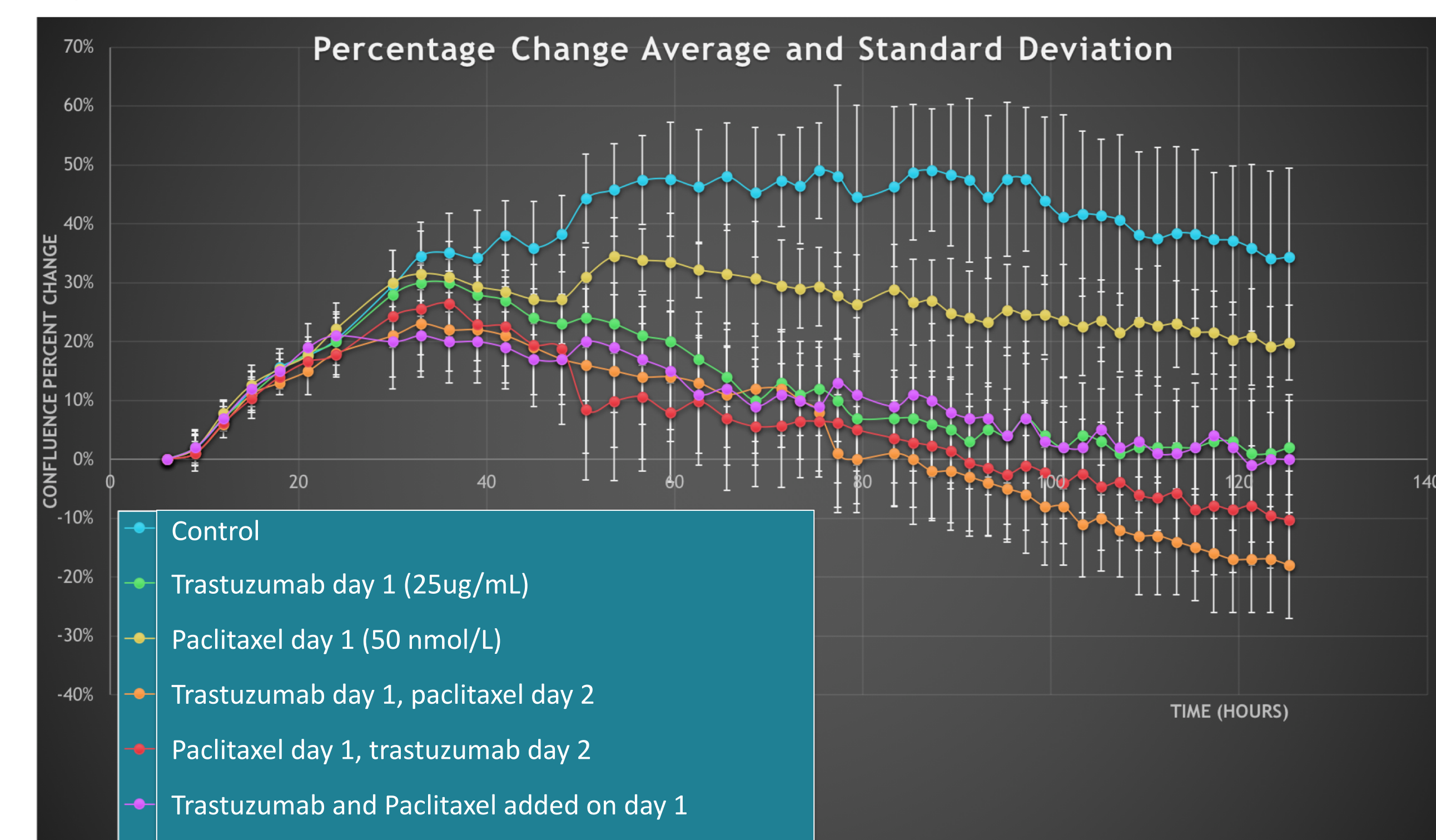
Data Type	Description
Tumor volume	Two separate sets recorded using caliper measurements <sup>1,2</sup>
Fraction of well-vascularized tumor	DCE-MRI data <sup>1</sup>
Fraction of necrosis in tumor	H&E stained histology data <sup>1</sup>
Fraction of hypoxia in tumor	FMISO-PET imaging data <sup>2</sup>
Fraction of immune response in tumor	Immunofluorescent stained histology data <sup>3</sup>



## In vitro data

- BT474 human derived breast cancer *in vitro* cell data measured by time-resolved microscopy
- Cultures treated with trastuzumab and/or paclitaxel on days 1 and 2 after cells are plated
- Change in confluence recorded over the course of seven days
- Trastuzumab dosing prior to paclitaxel potentially results in the greatest reduction in tumor cells

Figure 4: Preliminary *in vitro* results for cell culture data



## Formulation for Combining *in vivo* and *in vitro* scales

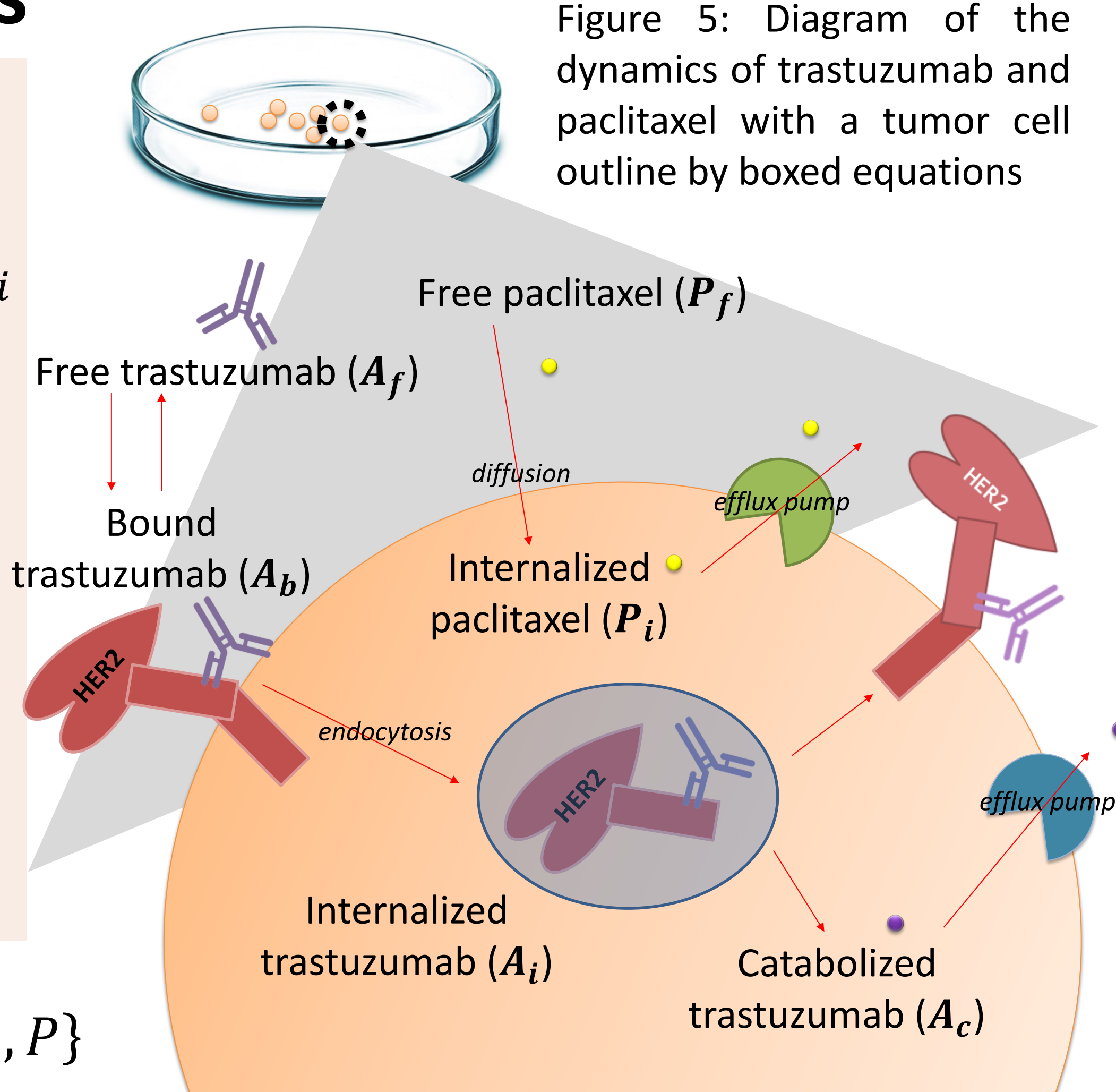
$$\frac{dT}{dt} = g(1 + \rho H) \left(1 - \frac{T}{K}\right) - dT - \mu_C T I$$

where  $d = f_d(A_b, P_i, g)$   
and  $g = f_g(A_b)$

- Trastuzumab-HER2 binding decreases proliferation and causes signaling cascades that can lead to cell death
- Paclitaxel arrests proliferation during mitosis leading to cell death (and therefore its effectiveness depends on cellular proliferation rates)
- Drug efflux will depend on a threshold amount ( $\gamma$ ) of the remaining previous drug dose

$$\begin{aligned} \frac{dA_f}{dt} &= -\beta_A A_f T + v_A A_b \\ \frac{dA_b}{dt} &= \beta_A A_f T - v_A A_b - \alpha_A A_b + \rho_A A_i \\ \frac{dA_i}{dt} &= \alpha_A A_b - \chi_A A_i - \rho_A A_i \\ \frac{dA_c}{dt} &= \chi_A A_i - f_\epsilon(\epsilon_A) A_c \\ \frac{dP_f}{dt} &= -\alpha_P P_f T + f_\epsilon(\epsilon_P) P_i \\ \frac{dP_i}{dt} &= \alpha_P P_f T - f_\epsilon(\epsilon_P) P_i \\ f_\epsilon(\epsilon_Y) &= \begin{cases} \epsilon_Y, & Z < \gamma \\ 0, & Z \geq \gamma \end{cases} \text{ where } Y, Z \in \{A, P\} \end{aligned}$$

Figure 5: Diagram of the dynamics of trastuzumab and paclitaxel with a tumor cell outline by boxed equations



## Next Steps

- Formulation of growth and death terms of the multiscale model
- Collect data for BT474 cells quantifying:
  - Internalized paclitaxel
  - Bound trastuzumab
  - Internalized trastuzumab
  - Catabolized trastuzumab
- Calibrate the multiscale model with *in vitro* data and make combination therapy predictions

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### References:

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- Sorace, A.G., et al., Mol Imaging Biol, 2017. **19**(1): p. 130-137
- Jarrett, A.M., et al., Under Review